

Synthesis and self-assembly of amphiphilic zinc chlorins possessing a 3¹-hydroxy group

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Abstract—Amphiphilic zinc chlorins possessing a 3¹-hydroxy group were prepared. Non-ionic (oligo)oxyethylene group, anionic sulfonate and cationic quaternary ammonium groups were introduced on the 17-position of a zinc chlorin moiety. All the three kinds of amphiphilic chlorophyll derivatives self-assembled in an aqueous medium to favor a formation of the anti-parallel dimeric structure. In contrast, the amphiphilic zinc chlorins formed large aggregates in non-polar organic solvent. These studies showed self-assemblies of amphiphilic zinc chlorin chromophores were controlled in environments. © 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

Generally, light-harvesting (LH) antenna of photosynthetic organisms consist of chlorophyll–protein complexes, in which antenna chlorophyll molecules are adequately arranged for effective energy transfer within a protein scaffold.¹ Green photosynthetic bacteria, however, have unique LH systems called chlorosomes, in which a number of bacteriochlorophyll (BChl)-*c*, *d* and *e* molecules (Fig. 1) self-aggregate to give rod-like oligomers.^{2–4} The light energy absorbed at the aggregated BChls is efficiently transferred into a baseplate of BChl-*a*–protein complex, indicating that the chlorosomal aggregate acts as a supramolecular light-harvesting device without any participation of proteins.⁵ BChls-*c*, *d* and *e* are characteristic in possessing the 3¹-hydroxy group, which plays important roles in the self-assembly. The C=O···H–O···Mg bonding among the 13¹-keto and 3¹-hydroxy groups and central magnesium is found in the self-aggregate.⁶ Interestingly, BChl molecules isolated from natural chlorosomes self-aggregate in vitro to reproduce the supramolecular aggregates.⁷ The strong excitonic interaction among BChl molecules results in red-shifted absorption bands and the intense CD signals in both in vivo and in vitro aggregates.^{7–9} Natural chlorosomal BChls have a hydrophobic large alkyl chain such as farnesyl, stearyl, phytol etc. at the 17⁴-position.¹⁰ The lipophilic BChls are organized within a

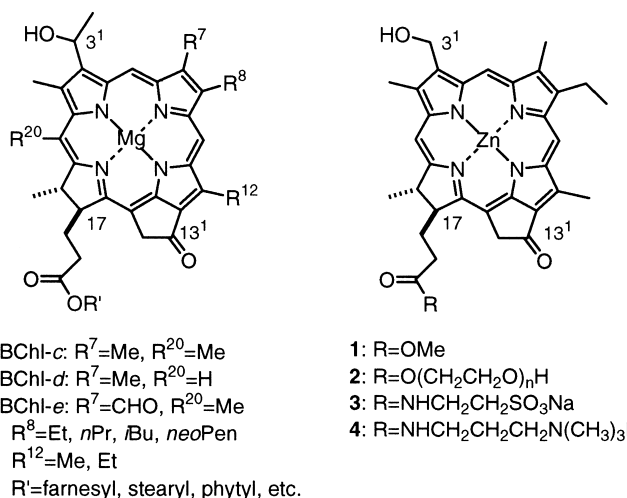


Figure 1. Structures of chlorosomal BChls and synthetic zinc chlorins 1–4.

hydrophobic region by monolayer of galactolipids. Although the precise suprastructure of the rod-like chlorosomal aggregate is not clarified, one of the proposed models suggested that the alkyl chain at the 17⁴-position is oriented to outside the rod-structure.³

The synthetic zinc chlorins with 3¹-hydroxy group such as **1** (Fig. 1) similarly form chlorosome-type aggregates in vitro.¹¹ Moreover, the artificial model of chlorosomal aggregate acts as light-harvesting device as well as natural BChls-*c*, *d* and *e* do in a chlorosome.^{12,13} The synthetic chlorophyll derivative is relatively stable compared to the natural pigments and the substituents are readily modified to

Keywords: amphiphilic molecule; chlorophyll; photosynthesis; self-aggregate.

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produce novel synthetic chlorophylls.^{14,15} The synthetic zinc chlorin **1** is monomeric in a polar organic solvent, such as methanol, tetrahydrofuran (THF), pyridine, etc. When the monomeric solution was diluted with an excess amount of non-polar organic solvent¹¹ or water,¹³ zinc chlorin **1** immediately self-aggregated to form their precipitates in the new environment. In contrast, the zinc chlorin **1** formed stable aqueous aggregates within a microheterogeneous medium produced by the added surfactants such as phosphatidyl choline or Triton X-100.^{13,16}

Self-assembly of the amphiphilic molecules in an aqueous medium gives unique aggregates with a micellar, hexagonal or lamellar suprastructure. The structure of the hydrophilic and hydrophobic parts in the amphiphilic molecule strongly affects the suprastructure. The (metallo)porphyrin possessing hydrophilic groups self-aggregates to give fibriform suprastructure in an aqueous medium, in which strong exciton coupling among porphyrin chromophore was observed.¹⁷ On the other hand, a free-base pyropheophorbide-*a* derivative possessing a cationic ammonium group formed dimeric aggregate in an aqueous medium.¹⁸ The unique supramolecular architecture of the aggregated amphiphilic tetrapyrroles was probably induced by the strong π - π interaction between the tetrapyrrole macrocycles.

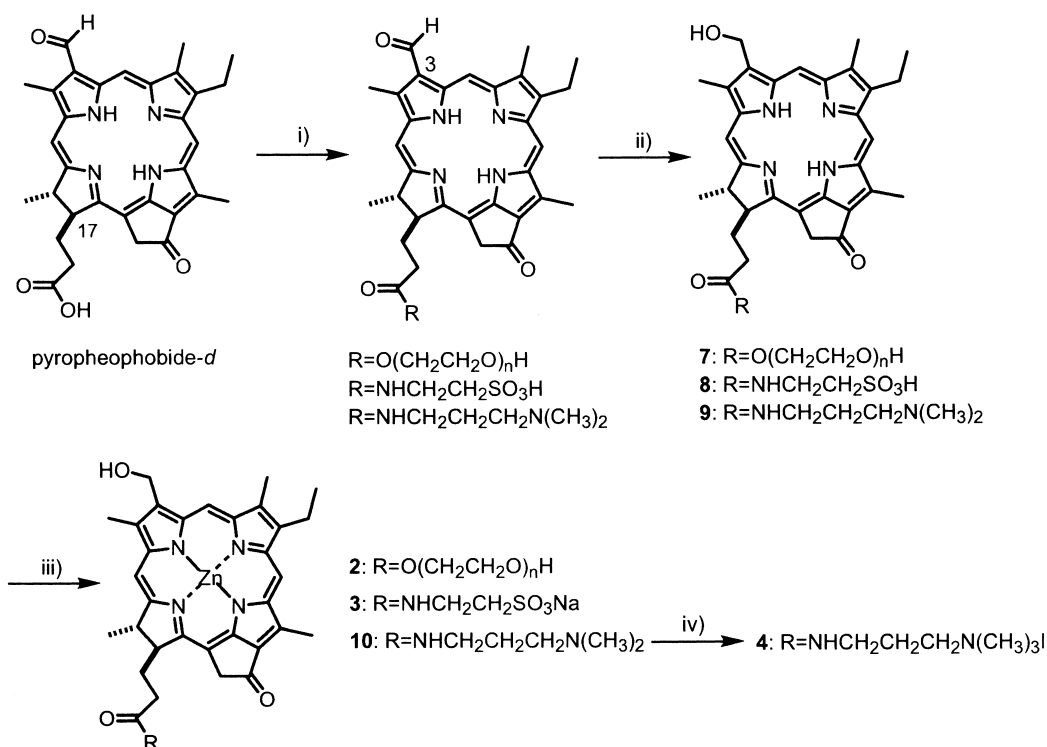
Here, we report that syntheses and self-assemblies of novel amphiphilic zinc analogues of chlorosomal BCChs possessing a hydrophilic group. The cationic, anionic and non-ionic hydrophilic groups were introduced on the 17-substituent of zinc 3¹-hydroxy-chlorins. The amphiphilic pigments self-assembled to form dimeric or large aggregates in aqueous or non-polar organic media, respectively.

2. Results and discussion

2.1. Synthesis of amphiphilic zinc chlorins possessing 3¹-hydroxy group

The non-ionic hydrophilic groups were introduced into chlorins as follows (Scheme 1). The propionic acid chain at the 17-position of pyropheophorbide-*d* was esterified with glycol by using 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC·HCl) and 4-(*N,N*-dimethylamino)pyridine (DMAP).¹⁹ Mono- (a), di- (b), tri- (c) and tetraethylene glycol (d), and polyethylene glycol (PEG) 300 (e), 400 (f), 600 (g), 1000 (h), 1540 (i) and 2000 (j) were used for the esterification. The average number of oxyethylene group of PEG 300, 400, 600, 1000, 1540 and 2000 (*n* in Scheme 1) were 6.4, 8.7, 13.3, 22.6, 34.0 and 44.1, respectively. The 3-formyl group of the esterified chlorin was selectively reduced into the 3-hydroxymethyl group as in **7** and the following zinc metallation¹¹ gave corresponding zinc chlorins **2** containing monoesterified (oligo)ethylene glycols and PEGs, 17²-COO(CH₂CH₂O)_{*n*}H; **2a** (*n*=1), **2b** (2), **2c** (3), **2d** (4), **2e** (6.4), **2f** (8.7), **2g** (13.3), **2h** (22.6), **2i** (34.0) and **2j** (44.1).

The anionic zinc chlorin **3** (sodium salt form) was prepared by condensation of pyropheophorbide-*d* with 2-aminoethanesulfonic acid (taurine),¹⁹ followed by reduction of 3-formyl group (**8**), zinc metallation and neutralization. The cationic chlorin possessing a quaternary ammonium group was prepared as follows. Pyropheophorbide-*d* was reacted with *N,N*-dimethyl-1,3-propanediamine to give the corresponding amide. After reduction of 3-formyl group, the free-base chlorin **9** was zinc-metallated. The tertiary amino group of **10** was methylated to the quaternary ammonium



Scheme 1. Synthesis of amphiphilic zinc 3-hydroxymethyl-chlorins **2–4**. (i) HO(CH₂CH₂O)_{*n*}H, H₂NCH₂CH₂SO₃H or H₂NCH₂CH₂CH₂N(CH₃)₂, EDC·HCl, DMAP/CH₂Cl₂, (ii) *t*BuNH₂·BH₃/CH₂Cl₂, (iii) Zn(OAc)₂·2H₂O/MeOH, CH₂Cl₂ and aq. NaHCO₃, (iv) MeI/MeOH, CH₂Cl₂.

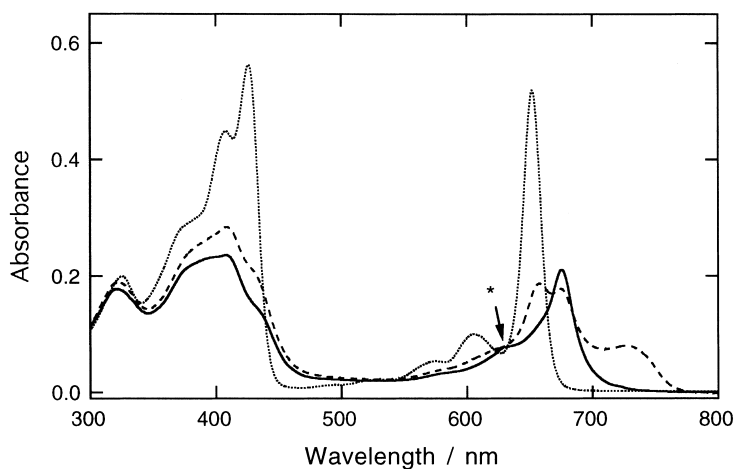


Figure 2. Visible absorption spectra of non-ionic zinc chlorin **2g** (ca. 10 μM); \cdots in methanol, $---$ in 1% MeOH–water immediately after aggregation (20 min), $—$ in 1% MeOH–water after standing for 2 days.

group by the addition of methyl iodide to give cationic zinc chlorin **4** (iodide form). The solubility of the amphiphilic zinc chlorins **2–4** in an aqueous media was dependent upon the structure of the hydrophilic group. Ionic zinc chlorins (**3**, **4**) and some non-ionic zinc chlorins **2g–j** ($n \geq 13.3$) were well soluble in water. In contrast, the other non-ionic zinc chlorins **2a–f** possessing a shorter (oligo)oxyethylene group (≤ 8.7) were sparingly soluble in water.

2.2. Amphiphilic zinc chlorins in an aqueous medium

Zinc chlorin **2g** possessing PEG 600 ($n=13.3$) gave absorption maxima at 425 and 652 nm in methanol showing the zinc chlorin as its monomeric form (the dotted line of Fig. 2). The other amphiphilic zinc chlorins **2–4** also gave similar visible spectra, indicating that a hydrophilic part of the 17²-position did not affect the optical properties of monomeric zinc chlorin. When the methanol solution of **2g** was diluted with 99-fold volume of water, the absorption spectrum was red-shifted due to the self-aggregation. Non-ionic zinc chlorin **2g** gave Qy bands at 652, 675 and 730 nm in 1% MeOH–H₂O (the dashed line of Fig. 2). Compared to the previous studies on aggregation of zinc chlorins,^{11,15} the absorption bands were ascribable to monomer, dimer and large aggregate of **2g**, respectively. The spectrum was, however, gradually changed at room temperature. After standing for 2 days, both the 652- and 730-nm bands decreased concomitantly with increase in 675-nm band (the solid line of Fig. 2). This result suggested that both the monomeric and large aggregate species of **2g** were relatively unstable and changed to stable dimeric species. That is, the hydrophilic polyoxyethylene (POE) chain disturbed the formation of a large aggregate. In addition, the slight increase in a 640-nm band (* on Fig. 2) was accompanied by dimerization of **2g**, which might be due to the other vibrational transition of dimer.

The other non-ionic zinc chlorins also gave the stable dimer in an aqueous medium. Fig. 3 shows the absorption spectra of zinc chlorins **2a–j** in 1% MeOH–water after standing for 3 days at room temperature. In zinc chlorins **2a–h** ($n \leq 22.6$), the 675 nm band of dimer was preferentially observed at the Qy region. Although the absorption band around 720 nm of large aggregates was slightly observed in

2a–d ($n \leq 4$), the red-shifted band gradually decreased during standing the solution. The zinc chlorins **2i** and **2j** with a longer POE chain ($n \geq 34.0$) showed the 657 nm band concomitant with a small band at 675 nm. The 675 nm band was enhanced with an increase of the concentration of zinc chlorin, aqueous 640 μM solution of **2j** gave the dimer as a major component. These results indicated that the non-ionic zinc chlorin **2** preferentially formed the dimeric structure in an aqueous medium, regardless of the length of POE chain.

The zinc chlorins possessing a long POE chain **2g–j** ($n \geq 13.3$) were soluble in water and showed an absorption maximum at 675 nm without addition of methanol. Thus, the presence of methanol was independent upon the formation of dimer. Similarly, both the anionic and cationic zinc chlorins **3** and **4** gave an absorption maximum at 675 nm in 1% MeOH–water (Fig. 4) and in only water.

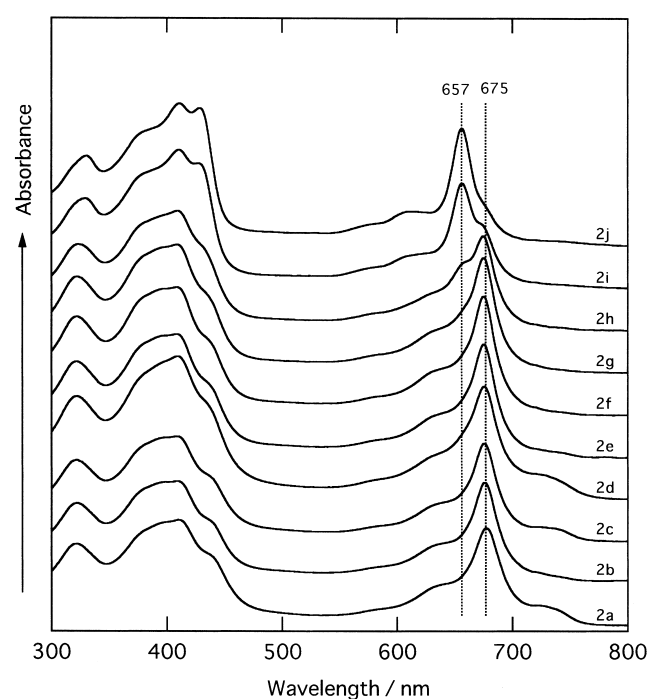


Figure 3. Visible absorption spectra of non-ionic zinc chlorins **2a–j** (ca. 10 μM) in 1% MeOH–water after standing for 3 days.

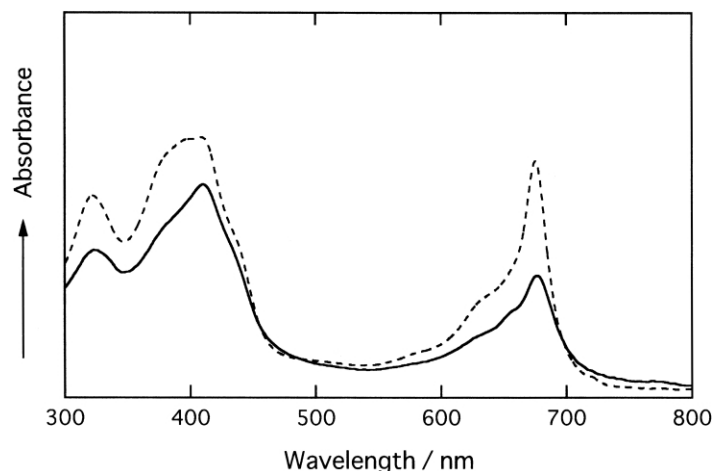


Figure 4. Visible absorption spectra of ionic zinc chlorins **3** and **4** (ca. 10 μ M) in 1% MeOH–water after standing for 3 days; - - - **3**, — **4**.

These results indicated that zinc chlorins possessing a hydrophilic group at the 17²-position self-aggregate to give the dimeric species in an aqueous medium, regardless of the structure of hydrophilic part.

Fluorescence emission and fluorescence excitation spectra of the non-ionic zinc chlorins **2** are measured as shown in Fig. 5. The emission spectrum of dimeric aggregates of amphiphilic zinc chlorin **2g** (the solid line of Fig. 5) gave an emission peak at 666-nm and a broad band around 700–750 nm, which was independent of excitation wavelength. The fluorescence excitation spectra measured at 710-nm (the dashed line of Fig. 5) was identical with absorption spectrum of the monomeric zinc chlorin measured in methanol. The spectral features showed that the dimeric zinc chlorin species was non- or much less fluorescent. Similar phenomena was reported in the other dimeric species of chlorophyll derivatives.^{18,20}

2.3. Suprastructure of dimeric species of amphiphilic zinc chlorins

The amphiphilic zinc chlorins possessing a 3¹-hydroxy group favored formation of dimeric species in an aqueous

medium, which was accompanied by a red-shift of the Qy band from 652 to 675 nm. Fig. 6A showed visible absorption spectra of amphiphilic zinc chlorin **5** lacking the 3¹-hydroxy group in an aqueous medium. Zinc 3-ethylchlorin **5** showed absorption maxima at 424, 647 nm in methanol (the dotted line of Fig. 6A) which were slightly red-shifted to 425, 652 nm in 1% methanol–water (the solid line of Fig. 6A). The small spectral changes were considered to solvent effect, thus **5** was monomeric in an aqueous medium. On the other hand, the free-base cationic 3¹-hydroxy-chlorin **6** showed absorption maxima at 406 and 660 nm in methanol, and 376 and 666 nm in an aqueous medium. The blue shift in Soret band ($\Delta\lambda=30$ nm) suggested that the free-base chlorins aggregated in an aqueous medium. Similar spectral feature was reported in another dimerization of the cationic pyropheophorbide-*a* derivatives in an aqueous medium.¹⁸ The π – π interaction between the chlorin chromophores should induce the dimerization of free-base **6** without any assistance of the 3¹-hydroxy groups. In the aqueous zinc chlorin **5**, the surrounding methanol or water might coordinate to central zinc. The solvation disturbed the π – π interaction, which resulted in formation of monomeric zinc chlorin **5** in an aqueous medium. These spectral studies showed that both

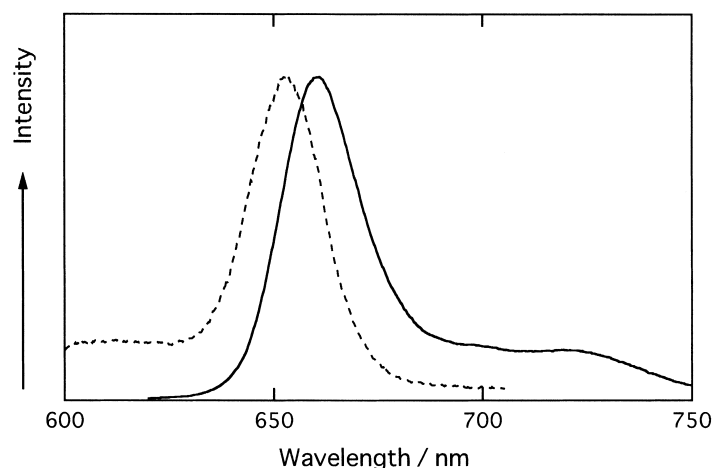


Figure 5. Fluorescence emission (excited at 450 nm: —) and fluorescence excitation (measured at 710 nm: - - -) of non-ionic zinc chlorin **2g** (ca. 10 μ M) in 1% MeOH–water after standing for 3 days.

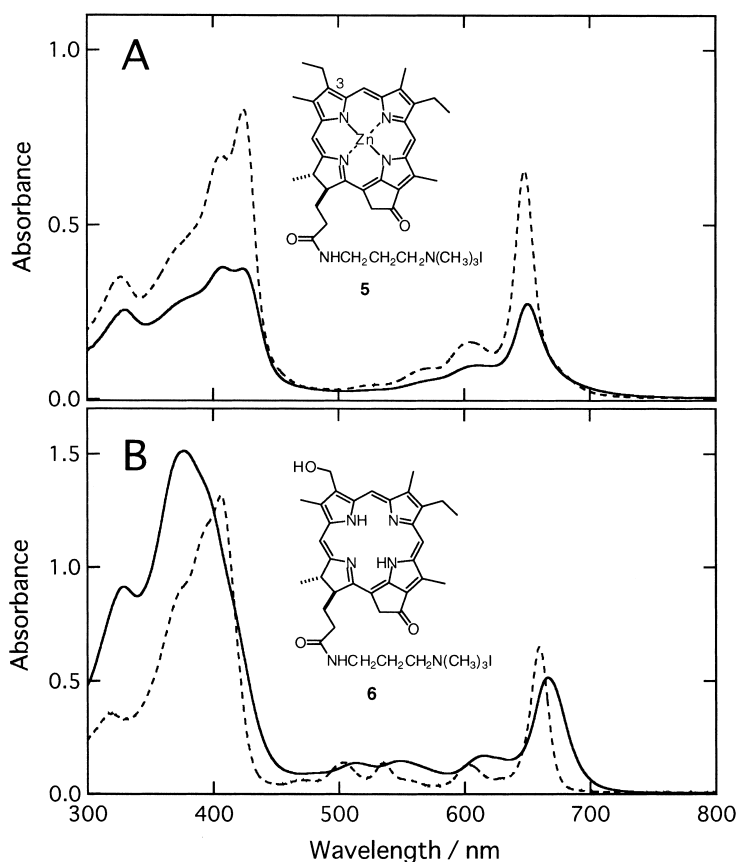


Figure 6. Visible absorption spectra of zinc 3-ethyl-chlorin **5** (A) and free-base chlorin **6** (B) in MeOH (---) and in 1% MeOH–water (—).

the central zinc and 3¹-hydroxy group were necessary for formation of the dimeric species characterized by 675 nm absorption band shown in Figs. 2–4.

As a result, the oxygen atom of the 3¹-hydroxy group was coordinated to the central zinc in the dimer of 2–4. In addition, the resonance Raman spectrum of the dimeric aggregate of zinc chlorin **2i** showed maxima at 1563 and 1621 cm⁻¹. The two peaks were assigned to C–C or C–N stretchings of five coordinated zinc chlorin moiety.⁶ The two structural models are available for the dimeric structure of amphiphilic zinc chlorins, the parallel structure I and the anti-parallel structure II (Fig. 7). The visible absorption spectra of the mixed aggregate of zinc 3¹-hydroxy-chlorin **4** and zinc 3-ethyl-chlorin **5** in an aqueous medium was similar to the linear combination of that of pure **4** and **5** (data not shown). Thus, the dimerization of amphiphilic zinc 3¹-hydroxy-chlorin **4** was not disturbed by addition of **5**. This result indicated that the anti-parallel structure II was favorable for dimeric species of the amphiphilic zinc

chlorins, in which the 3¹-hydroxy group of both zinc chlorins were coordinated to the central zinc each other.

2.4. Amphiphilic zinc chlorins in a non-polar organic solvent

The amphiphilic zinc chlorins 2–4 were dissolved in a small amount of tetrahydrofuran (THF) and diluted with a large volume of hexane. The Qy absorption maximum of the non-ionic zinc chlorin **2g** showed a red-shifted and broadened band at around 740 nm with a 710-nm shoulder, which was not shifted after standing for a long period (the solid line of Fig. 8). In addition, the precipitate of the aggregated **2g** was immediately appeared in hexane solution. Similar results were observed in the other non-ionic zinc chlorins **2a–j** in 1% THF–hexane. These results indicated that the non-ionic **2** did not form dimeric species but several large aggregates in a non-polar organic medium. The hydrophilic oxyethylene group probably oriented to inside the large aggregate. On the other hand, in the anionic and cationic

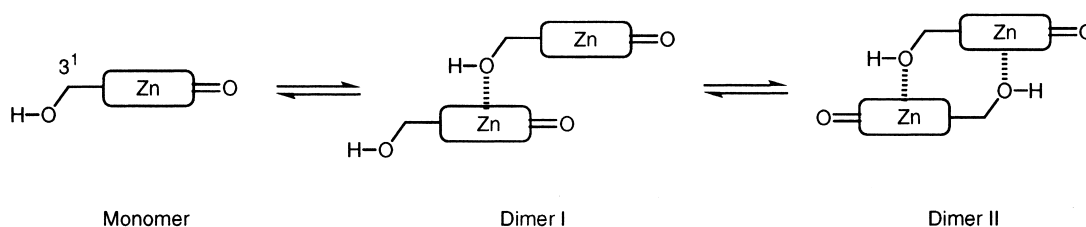


Figure 7. Proposed suprastructures of dimeric aggregate of amphiphilic zinc chlorin.

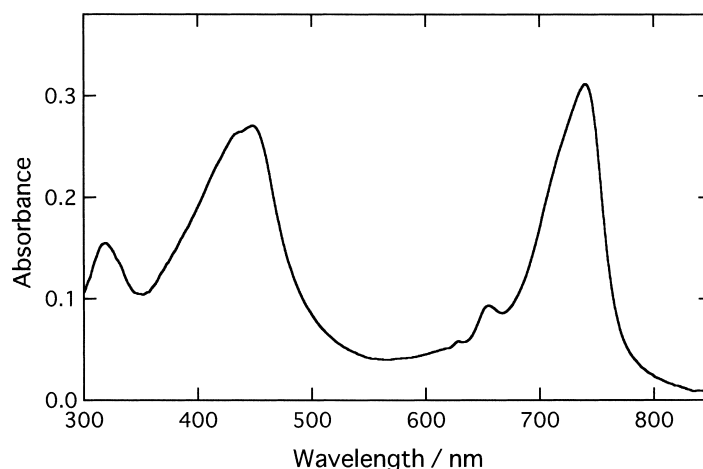


Figure 8. Visible absorption spectrum of non-ionic zinc chlorin **2g** (ca. 10 μ M) in 1% THF–hexane.

zinc chlorins **3** and **4**, the formation of precipitate was too rapid to measure the absorption spectrum.

2.5. Self-assembly processes of amphiphilic zinc chlorins

In an aqueous medium, amphiphilic zinc chlorins formed a large aggregate immediately after dilution with water. The previous studies showed the hydrophobic zinc chlorin **1** self-aggregate to give precipitate in an aqueous media.¹³ Thus, the aggregation of the hydrophobic zinc chlorin continuously proceeded until the precipitate was formed. In contrast, the large aggregate of the amphiphilic zinc chlorin was formed without any precipitate, indicating that the hydrophilic group of zinc chlorin disturbed the further aggregation. The hydrophilic group at the 17²-position should be oriented to the bulk water to surround the core aggregated zinc chlorin moieties. In a non-polar medium, the hydrophilic part of the amphiphilic zinc chlorins, however, did not cover the aggregated zinc chlorins, which resulted in formation of precipitate in hexane.

The temporarily formed micelle-like aggregate in an aqueous media was, however, thermodynamically unstable, thus it gradually disaggregated to the dimeric species. The suprastructure of the aqueous assembly of amphiphilic molecules is, in general, affected by the structures of hydrophilic and hydrophobic parts. However, in all the amphiphilic zinc chlorins **2–4** examined here, the anti-parallel dimeric species should be the most favorable structure in aqueous self-assemblies.

3. Conclusion

Zinc 3¹-hydroxy-chlorins possessing a hydrophilic part on the 17-position were prepared. In an aqueous medium, the amphiphilic zinc chlorins formed a large aggregate as a kinetic product, which was thermodynamically unstable to change more favorable dimeric species with anti-parallel structure. In contrast, the amphiphilic zinc chlorins formed large aggregates in a non-polar organic medium. Thus, environments strongly affect the suprastructures of amphiphilic zinc chlorin aggregates; water molecules as a solvent

interacted with the hydrophilic group to stabilize a compact dimer and hexane molecules repulsed them to make large self-aggregates through interaction among the hydrophilic group in a supramolecule.

4. Experimental

4.1. Apparatus

Visible absorption and fluorescence spectra were measured by Shimadzu UV-3100 spectrophotometer and Hitachi F-4500 fluorescence spectrophotometer, respectively. Resonance Raman spectra were measured with Jasco NR-1800 spectrophotometer using a 457.9 nm line of Ar⁺ laser. ¹H NMR spectra were recorded with Bruker DPX-400 spectrophotometer with tetramethylsilane as internal standard. Fast atomic bombardment (FAB)-mass spectra (MS) were measured with a JEOL GCmate II spectrometer. FAB-MS samples were dissolved in CH₂Cl₂ and 3-nitrobenzyl alcohol or glycerol was used as a matrix. High performance liquid chromatography (HPLC) was performed with a Shimadzu LC-10AS pump and an SPD-M10AV diode array detector.

4.2. Materials

Pyropheophorbide-*d* was prepared according to the reported procedure.¹² Polyethylene glycol 300, 400, 600, 1000, 1540 and 2000 were purchased from Katayama Chemical Co. Ltd. (Japan). The average molecular weights of the PEG samples are 300, 402, 603, 1011, 1514, 2047, respectively. Flash column chromatography (FCC) was performed with silica gel (Merck, Kieselgel 60, 9385). HPLC was done with a packed ODS column (Gelpack GL-OP100, Hitachi Chemical Co., (Japan), 6.0 ϕ ×150 mm or Cosmosil 5C₁₈-ARII, Nacalai tesque, (Japan), 6.0 ϕ ×250 mm).

4.3. General procedures

Reduction of the 3-formyl group of a chlorin to the 3-hydroxymethyl group and zinc metallation of a chlorin were performed according to the reported procedures.¹¹

Condensation of pyropheophorbide-*d* with glycol or amine. Pyropheophorbide-*d* (0.025 mmol) was dissolved with dried dichloromethane (20 mL). EDC·HCl (0.15 mmol) and DMAP (0.3 mmol) were added to the solution with stirring at 0°C. The excess amount of glycol or amine (>0.5 mmol) was added to the solution and stirred for 5 h. The reaction mixture was poured into 2% HCl and CH₂Cl₂. The organic layer was separated and washed with 4% NaHCO₃, water and dried with Na₂SO₄. The product was purified with FCC or HPLC to give the corresponding glycol monoester or amide.

***N*-Methylation of tertiary amine.** To a dry dichloromethane solution (3.0 mL) of tertiary amine compound (0.02 mmol) was added dry methanol (0.5 mL) and dry methyl iodide (0.5 mL). The reaction mixture was stirred for 20 h at room temperature under N₂ in the dark. After evaporation of the solvent, the residue was purified with HPLC to give the corresponding ammonium iodide.

4.4. Spectral data

4.4.1. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* ethylene glycol monoester (7a). Esterification of pyropheophorbide-*d* with ethylene glycol followed by reduction of 3-formyl group gave black solids of **7a**, 72% yield (FCC 2% MeOH–CH₂Cl₂). Mp 226–229°C; VIS (CH₂Cl₂) λ_{max} (relative intensity)=662 (0.46), 606 (0.07), 536 (0.09), 505 (0.09), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.41, 9.39, 8.53 (each 1H, s, 5-, 10-, 20-H), 5.87 (2H, s, 3-CH₂), 5.17, 5.04 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.43 (1H, dq, *J*=2, 7 Hz, 18-H), 4.24 (1H, dt, *J*=8, 2 Hz, 17-H), 4.02–4.13 (2H, m, COOCH₂), 3.65 (2H, q, *J*=7 Hz, 8-CH₂), 3.58–3.64 (2H, m, COOCH₂CH₂), 3.57, 3.40, 3.24 (each 3H, s, 2-, 7-, 12-CH₃), 2.47–2.65, 2.17–2.37 (each 2H, m, 17-CH₂CH₂), 1.77 (3H, d, *J*=7 Hz, 18-CH₃), 1.67 (3H, t, *J*=7 Hz, 8¹-CH₃), 0.20, –1.86 (each 1H, s, NH). MS (FAB) found: *m/z*=583. Calcd for C₃₄H₃₉N₄O₅: MH⁺, 583.

4.4.2. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* diethylene glycol monoester (7b). Esterification of pyropheophorbide-*d* with diethylene glycol followed by reduction of 3-formyl group gave black solids of **7b**, 71% yield (FCC 2–3% MeOH–CH₂Cl₂). Mp 122–125°C; VIS (CH₂Cl₂) λ_{max}=662 (0.47), 606 (0.08), 536 (0.09), 506 (0.09), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.48, 9.44, 8.55 (each 1H, s, 5-, 10-, 20-H), 5.90 (2H, s, 3-CH₂), 5.25, 5.09 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.48 (1H, dq, *J*=2, 7 Hz, 18-H), 4.28 (1H, dt, *J*=8, 2 Hz, 17-H), 4.08–4.19 (2H, m, COOCH₂), 3.69 (2H, q, *J*=7 Hz, 8-CH₂), 3.43–3.67 (6H, m, COOCH₂CH₂OCH₂CH₂OH), 3.65, 3.42, 3.26 (each 3H, s, 2-, 7-, 12-CH₃), 2.52–2.74, 2.20–2.38 (each 2H, m, 17-CH₂CH₂), 1.80 (3H, d, *J*=7 Hz, 18-CH₃), 1.69 (3H, t, *J*=7 Hz, 8¹-CH₃), 0.32, –1.77 (each 1H, s, NH). MS (FAB) found: *m/z*=627. Calcd for C₃₆H₄₃N₄O₆: MH⁺, 627.

4.4.3. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* triethylene glycol monoester (7c). Esterification of pyropheophorbide-*d* with triethylene glycol followed by reduction of 3-formyl group gave black solids of **7c**, 75% yield (FCC 4% MeOH–CH₂Cl₂). Mp 97–99°C; VIS (CH₂Cl₂) λ_{max}=662 (0.47), 606 (0.07), 536 (0.09), 504 (0.09), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.53, 9.47, 8.57

(each 1H, s, 5-, 10-, 20-H), 5.87–5.97 (2H, m, 3-CH₂), 5.27, 5.12 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.49 (1H, dq, *J*=2, 7 Hz, 18-H), 4.33 (1H, dt, *J*=8, 2 Hz, 17-H), 3.98–4.12 (2H, m, COOCH₂), 3.71 (2H, q, *J*=7 Hz, 8-CH₂), 3.68, 3.43, 3.27 (each 3H, s, 2-, 7-, 12-CH₃), 3.32–3.55 (10H, m, COOCH₂CH₂O(CH₂CH₂O)₂), 2.63–2.74, 2.46–2.57, 2.16–2.40 (1H+1H+2H, m, 17-CH₂CH₂), 1.80 (3H, d, *J*=7 Hz, 18-CH₃), 1.70 (3H, t, *J*=7 Hz, 8¹-CH₃), 0.34, –1.76 (each 1H, s, NH). MS (FAB) found: *m/z*=671. Calcd for C₃₈H₄₇N₄O₇: MH⁺, 671.

4.4.4. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* tetraethylene glycol monoester (7d). Esterification of pyropheophorbide-*d* with tetraethylene glycol followed by reduction of 3-formyl group gave black solids of **7d**, 69% yield (FCC 5% MeOH–CH₂Cl₂). Mp 89–92°C; VIS (CH₂Cl₂) λ_{max}=663 (0.47), 606 (0.08), 536 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.44, 9.41, 8.55 (each 1H, s, 5-, 10-, 20-H), 5.89, 5.84 (each 1H, d, *J*=13 Hz, 3-CH₂), 5.21, 5.07 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.46 (1H, dq, *J*=2, 7 Hz, 18-H), 4.28 (1H, dt, *J*=8, 2 Hz, 17-H), 4.03–4.11, 3.93–4.00 (each 1H, m, COOCH₂), 3.66 (2H, q, *J*=7 Hz, 8-CH₂), 3.62, 3.43, 3.24 (each 3H, s, 2-, 7-, 12-CH₃), 3.32–3.59 (14H, m, COOCH₂CH₂O(CH₂CH₂O)₃), 2.43–2.74, 2.11–2.34 (each 2H, m, 17-CH₂CH₂), 1.78 (3H, d, *J*=7 Hz, 18-CH₃), 1.68 (3H, t, *J*=7 Hz, 8¹-CH₃), 0.23, –1.84 (each 1H, s, NH). MS (FAB) found: *m/z*=715 (MH⁺). Calcd for C₄₀H₅₁N₄O₈: MH⁺, 715.

4.4.5. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 300 monoester (7e). Esterification of pyropheophorbide-*d* with polyethylene glycol 300 (*n*=6.4) followed by reduction of 3-formyl group gave black solids of **7e**, 72% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 5.0 min). Mp 60–69°C; VIS (CH₂Cl₂) λ_{max}=663 (0.44), 606 (0.07), 536 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.46, 9.44, 8.55 (each 1H, s, 5-, 10-, 20-H), 5.90, 5.86 (each 1H, d, *J*=14 Hz, 3-CH₂), 5.23, 5.09 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.47 (1H, dq, *J*=2, 7 Hz, 18-H), 4.29 (1H, dt, *J*=8, 2 Hz, 17-H), 4.00–4.19 (2H, m, COOCH₂), 3.68 (2H, q, *J*=7 Hz, 8-CH₂), 3.28–3.68 (24H, m, COOCH₂CH₂O(CH₂CH₂O)_{5.4}), 3.63, 3.41, 3.25 (each 3H, s, 2-, 7-, 12-CH₃), 2.45–2.72, 2.19–2.34 (each 2H, m, 17-CH₂CH₂), 1.79 (3H, d, *J*=7 Hz, 18-CH₃), 1.68 (3H, t, *J*=7 Hz, 8¹-CH₃), 0.26, –1.82 (each 1H, s, NH). MS (FAB) found: *m/z*=671, 715, 759, 803, 847, 891, 935, 979. Calcd for C₃₈H₄₇N₄O₇: MH⁺, 671 (*n*=3), C₄₀H₅₁N₄O₈: MH⁺, 715 (*n*=4), C₄₂H₅₅N₄O₉: MH⁺, 759 (*n*=5), C₄₄H₅₉N₄O₁₀: MH⁺, 803 (*n*=6), C₄₆H₆₃N₄O₁₁: MH⁺, 847 (*n*=7), C₄₈H₆₇N₄O₁₂: MH⁺, 891 (*n*=8), C₅₀H₇₁N₄O₁₃: MH⁺, 935 (*n*=9), C₅₂H₇₅N₄O₁₄: MH⁺, 979 (*n*=10).

4.4.6. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 400 monoester (7f). Esterification of pyropheophorbide-*d* with polyethylene glycol 400 (*n*=8.7) followed by reduction of 3-formyl group gave black solids of **7f**, 83% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 4.8 min). Mp 57–64°C; VIS (CH₂Cl₂) λ_{max}=663 (0.44), 606 (0.08), 536 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) δ=9.48, 9.45, 8.56 (each 1H, s, 5-, 10-, 20-H), 5.91, 5.87 (each 1H, d, *J*=13 Hz, 3-CH₂), 5.24, 5.10 (each 1H, d, *J*=20 Hz, 13²-H₂), 4.48 (1H,

dq, $J=2, 7$ Hz, 18-H), 4.30 (1H, dt, $J=8, 2$ Hz, 17-H), 4.00–4.17 (2H, m, COOCH₂), 3.69 (2H, q, $J=7$ Hz, 8-CH₂), 3.29–3.68 (33H, m, COOCH₂CH₂O(CH₂CH₂O)_{7.7}), 3.65, 3.42, 3.25 (each 3H, s, 2-, 7-, 12-CH₃), 2.48–2.72, 2.20–2.35 (each 2H, m, 17-CH₂CH₂), 1.80 (3H, d, $J=7$ Hz, 18-CH₃), 1.69 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.27, –1.80 (each 1H, s, NH). MS (FAB) found: $m/z=715, 759, 803, 847, 891, 935, 979, 1023, 1067$. Calcd for C₄₀H₅₁N₄O₈: MH⁺, 715 ($n=4$), C₄₂H₅₅N₄O₉: MH⁺, 759 ($n=5$), C₄₄H₅₉N₄O₁₀: MH⁺, 803 ($n=6$), C₄₆H₆₃N₄O₁₁: MH⁺, 847 ($n=7$), C₄₈H₆₇N₄O₁₂: MH⁺, 891 ($n=8$), C₅₀H₇₁N₄O₁₃: MH⁺, 935 ($n=9$), C₅₂H₇₅N₄O₁₄: MH⁺, 979 ($n=10$), C₅₄H₇₉N₄O₁₅: MH⁺, 1023 ($n=11$), C₅₆H₈₃N₄O₁₆: MH⁺, 1067 ($n=12$).

4.4.7. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 600 monoester (7g). Esterification of pyropheophorbide-*d* with polyethylene glycol 600 ($n=13.3$) followed by reduction of 3-formyl group gave black solids of **7g**, 76% yield (HPLC, Cosmosil with 10% H₂O/MeOH 1.0 mL/min, the retention time was 13.8 min). Mp 52–55°C; VIS (CH₂Cl₂) $\lambda_{\max}=662$ (0.47), 605 (0.08), 536 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) $\delta=9.52, 9.48, 8.57$ (each 1H, s, 5-, 10-, 20-H), 5.93, 5.90 (each 1H, d, $J=13$ Hz, 3-CH₂), 5.26, 5.12 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.49 (1H, dq, $J=2, 7$ Hz, 18-H), 4.32 (1H, dt, $J=8, 2$ Hz, 17-H), 4.03–4.16 (2H, m, COOCH₂), 3.71 (2H, q, $J=7$ Hz, 8-CH₂), 3.36–3.71 (51H, m, COOCH₂CH₂O(CH₂CH₂O)_{12.3}), 3.67, 3.43, 3.27 (each 3H, s, 2-, 7-, 12-CH₃), 2.51–2.75, 2.23–2.37 (each 2H, m, 17-CH₂CH₂), 1.81 (3H, d, $J=7$ Hz, 18-CH₃), 1.70 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.34, –1.76 (each 1H, s, NH). MS (FAB) found: $m/z=847, 891, 935, 979, 1023, 1067, 1111, 1155, 1199, 1243, 1287$. Calcd for C₄₆H₆₃N₄O₁₁: MH⁺, 847 ($n=7$), C₄₈H₆₇N₄O₁₂: MH⁺, 891 ($n=8$), C₅₀H₇₁N₄O₁₃: MH⁺, 935 ($n=9$), C₅₂H₇₅N₄O₁₄: MH⁺, 979 ($n=10$), C₅₄H₇₉N₄O₁₅: MH⁺, 1023 ($n=11$), C₅₆H₈₃N₄O₁₆: MH⁺, 1067 ($n=12$), C₅₈H₈₇N₄O₁₇: MH⁺, 1111 ($n=13$), C₆₀H₉₁N₄O₁₈: MH⁺, 1155 ($n=14$), C₆₂H₉₅N₄O₁₉: MH⁺, 1199 ($n=15$), C₆₄H₉₉N₄O₂₀: MH⁺, 1243 ($n=16$), C₆₆H₁₀₃N₄O₂₁: MH⁺, 1287 ($n=17$).

4.4.8. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 1000 monoester (7h). Esterification of pyropheophorbide-*d* with polyethylene glycol 1000 ($n=22.6$) followed by reduction of 3-formyl group gave black solids of **7h**, 76% yield (HPLC, Cosmosil with 10% H₂O/MeOH 1.0 mL/min, the retention time was 12.6 min). Mp <30°C; VIS (CH₂Cl₂) $\lambda_{\max}=662$ (0.48), 605 (0.08), 535 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) $\delta=9.48, 9.46, 8.56$ (each 1H, s, 5-, 10-, 20-H), 5.91, 5.88 (2H, d, $J=13$ Hz, 3-CH₂), 5.24, 5.10 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.48 (1H, dq, $J=2, 7$ Hz, 18-H), 4.30 (1H, dt, $J=8, 2$ Hz, 17-H), 4.03–4.17 (2H, m, COOCH₂), 3.38–3.73 (88H, m, COOCH₂CH₂O(CH₂CH₂O)_{21.6}), 3.69 (2H, q, $J=7$ Hz, 8-CH₂), 3.65, 3.42, 3.26 (each 3H, s, 2-, 7-, 12-CH₃), 2.63–2.73, 2.50–2.61, 2.23–2.34 (1H+1H+2H, m, 17-CH₂CH₂), 1.80 (3H, d, $J=7$ Hz, 18-CH₃), 1.69 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.30, –1.79 (each 1H, s, NH). MS (FAB) found: $m/z=1111, 1155, 1199, 1243, 1287, 1331, 1375, 1419, 1463, 1507, 1551, 1595, 1639, 1683, 1727, 1771, 1815, 1859$. Calcd for C₅₈H₈₇N₄O₁₇: MH⁺, 1111 ($n=13$), C₆₀H₉₁N₄O₁₈: MH⁺, 1155 ($n=14$), C₆₂H₉₅N₄O₁₉: MH⁺, 1199 ($n=15$), C₆₄H₉₉N₄O₂₀: MH⁺, 1243 ($n=16$),

C₆₆H₁₀₃N₄O₂₁: MH⁺, 1287 ($n=17$), C₆₈H₁₀₇N₄O₂₂: MH⁺, 1331 ($n=18$), C₇₀H₁₁₁N₄O₂₃: MH⁺, 1375 ($n=19$), C₇₂H₁₁₅N₄O₂₄: MH⁺, 1419 ($n=20$), C₇₄H₁₁₉N₄O₂₅: MH⁺, 1463 ($n=21$), C₇₆H₁₂₃N₄O₂₆: MH⁺, 1507 ($n=22$), C₇₈H₁₂₇N₄O₂₇: MH⁺, 1551 ($n=23$), C₈₀H₁₃₁N₄O₂₈: MH⁺, 1595 ($n=24$), C₈₂H₁₃₅N₄O₂₉: MH⁺, 1639 ($n=25$), C₈₄H₁₃₉N₄O₃₀: MH⁺, 1683 ($n=26$), C₈₆H₁₄₃N₄O₃₁: MH⁺, 1727 ($n=27$), C₈₈H₁₄₇N₄O₃₂: MH⁺, 1771 ($n=28$), C₉₀H₁₅₁N₄O₃₃: MH⁺, 1815 ($n=29$), C₉₂H₁₅₅N₄O₃₄: MH⁺, 1859 ($n=30$).

4.4.9. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 1540 monoester (7i). Esterification of pyropheophorbide-*d* with polyethylene glycol 1540 ($n=34.0$) followed by reduction of 3-formyl group gave black solids of **7i**, 72% yield (HPLC, Cosmosil with 10% H₂O/MeOH 1.0 mL/min, the retention time was 11.5 min). Mp 40–43°C; VIS (CH₂Cl₂) $\lambda_{\max}=662$ (0.46), 605 (0.08), 536 (0.09), 505 (0.10), 411 nm (1.00); ¹H NMR (CDCl₃) $\delta=9.51, 9.48, 8.57$ (each 1H, s, 5-, 10-, 20-H), 5.92, 5.89 (2H, d, $J=13$ Hz, 3-CH₂), 5.26, 5.11 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.49 (1H, dq, $J=2, 7$ Hz, 18-H), 4.31 (1H, dt, $J=8, 2$ Hz, 17-H), 4.03–4.17 (2H, m, COOCH₂), 3.39–3.73 (134H, m, COOCH₂CH₂O(CH₂CH₂O)₃₃), 3.70 (2H, q, $J=7$ Hz, 8-CH₂), 3.67, 3.43, 3.27 (each 3H, s, 2-, 7-, 12-CH₃), 2.64–2.74, 2.51–2.61, 2.23–2.36 (1H+1H+2H, m, 17-CH₂CH₂), 1.81 (3H, d, $J=7$ Hz, 18-CH₃), 1.70 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.33, –1.77 (each 1H, s, NH). MS (FAB) found: $m/z=1595, 1639, 1683, 1727, 1771, 1815, 1859, 1903, 1947, 1991, 2035, 2079, 2123, 2167, 2211, 2255, 2299, 2343, 2387$. Calcd for C₈₀H₁₃₁N₄O₂₈: MH⁺, 1595 ($n=24$), C₈₂H₁₃₅N₄O₂₉: MH⁺, 1639 ($n=25$), C₈₄H₁₃₉N₄O₃₀: MH⁺, 1683 ($n=26$), C₈₆H₁₄₃N₄O₃₁: MH⁺, 1727 ($n=27$), C₈₈H₁₄₇N₄O₃₂: MH⁺, 1771 ($n=28$), C₉₀H₁₅₁N₄O₃₃: MH⁺, 1815 ($n=29$), C₉₂H₁₅₅N₄O₃₄: MH⁺, 1859 ($n=30$), C₉₄H₁₅₉N₄O₃₅: MH⁺, 1903 ($n=31$), C₉₆H₁₆₃N₄O₃₆: MH⁺, 1947 ($n=32$), C₉₈H₁₆₇N₄O₃₇: MH⁺, 1991 ($n=33$), C₁₀₀H₁₇₁N₄O₃₈: MH⁺, 2035 ($n=34$), C₁₀₂H₁₇₅N₄O₃₉: MH⁺, 2079 ($n=35$), C₁₀₄H₁₇₉N₄O₄₀: MH⁺, 2123 ($n=36$), C₁₀₆H₁₈₃N₄O₄₁: MH⁺, 2167 ($n=37$), C₁₀₈H₁₈₇N₄O₄₂: MH⁺, 2211 ($n=38$), C₁₁₀H₁₉₁N₄O₄₃: MH⁺, 2255 ($n=39$), C₁₁₂H₁₉₅N₄O₄₄: MH⁺, 2299 ($n=40$), C₁₁₄H₁₉₉N₄O₄₅: MH⁺, 2343 ($n=41$), C₁₁₆H₂₀₃N₄O₄₆: MH⁺, 2387 ($n=42$).

4.4.10. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-*a* polyethylene glycol 2000 monoester (7j). Esterification of pyropheophorbide-*d* with polyethylene glycol 2000 ($n=44.1$) followed by reduction of 3-formyl group gave black solids of **7j**, 68% yield (HPLC, Cosmosil with 10% H₂O/MeOH 1.0 mL/min, the retention time was 10.7 min). Mp 46–49°C; VIS (CH₂Cl₂) $\lambda_{\max}=662$ (0.48), 605 (0.08), 536 (0.09), 505 (0.10), 410 nm (1.00); ¹H NMR (CDCl₃) $\delta=9.54, 9.49, 8.58$ (each 1H, s, 5-, 10-, 20-H), 5.92 (2H, s, 3-CH₂), 5.27, 5.12 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.50 (1H, dq, $J=2, 7$ Hz, 18-H), 4.33 (1H, dt, $J=8, 2$ Hz, 17-H), 4.03–4.17 (2H, m, COOCH₂), 3.39–3.83 (182H, m, COOCH₂-CH₂O(CH₂CH₂O)_{43.1}), 3.71 (2H, q, $J=7$ Hz, 8-CH₂), 3.69, 3.44, 3.28 (each 3H, s, 2-, 7-, 12-CH₃), 2.65–2.75, 2.52–2.62, 2.24–2.37 (1H+1H+2H, m, 17-CH₂CH₂), 1.81 (3H, d, $J=7$ Hz, 18-CH₃), 1.71 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.36, –1.75 (each 1H, s, NH). MS (FAB) found: $m/z=1947, 1991, 2035, 2079, 2123, 2167, 2211, 2255, 2299, 2343,$

2387, 2431, 2475, 2519, 2563, 2607, 2651, 2695, 2739, 2783, 2827. Calcd for $C_{96}H_{163}N_4O_{36}$: MH^+ , 1947 ($n=32$), $C_{98}H_{167}N_4O_{37}$: MH^+ , 1991 ($n=33$), $C_{100}H_{171}N_4O_{38}$: MH^+ , 2035 ($n=34$), $C_{102}H_{175}N_4O_{39}$: MH^+ , 2079 ($n=35$), $C_{104}H_{179}N_4O_{40}$: MH^+ , 2123 ($n=36$), $C_{106}H_{183}N_4O_{41}$: MH^+ , 2167 ($n=37$), $C_{108}H_{187}N_4O_{42}$: MH^+ , 2211 ($n=38$), $C_{110}H_{191}N_4O_{43}$: MH^+ , 2255 ($n=39$), $C_{112}H_{195}N_4O_{44}$: MH^+ , 2299 ($n=40$), $C_{114}H_{199}N_4O_{45}$: MH^+ , 2343 ($n=41$), $C_{116}H_{203}N_4O_{46}$: MH^+ , 2387 ($n=42$), $C_{118}H_{207}N_4O_{47}$: MH^+ , 2431 ($n=43$), $C_{120}H_{211}N_4O_{48}$: MH^+ , 2475 ($n=44$), $C_{122}H_{215}N_4O_{49}$: MH^+ , 2519 ($n=45$), $C_{124}H_{219}N_4O_{50}$: MH^+ , 2563 ($n=46$), $C_{126}H_{223}N_4O_{51}$: MH^+ , 2607 ($n=47$), $C_{128}H_{227}N_4O_{52}$: MH^+ , 2651 ($n=48$), $C_{130}H_{231}N_4O_{53}$: MH^+ , 2695 ($n=49$), $C_{132}H_{235}N_4O_{54}$: MH^+ , 2739 ($n=50$), $C_{134}H_{239}N_4O_{55}$: MH^+ , 2783 ($n=51$), $C_{136}H_{243}N_4O_{56}$: MH^+ , 2827 ($n=52$).

4.4.11. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a ethylene glycol monoester (2a). Zinc metallation of **7a** gave dark green solids of **2a**, 92% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 9.4 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 601 (0.10), 567 (0.05), 521 (0.03), 424 (1.00), 405 nm (0.55); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57$, 9.31, 8.35 (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.18, 5.07 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.40 (1H, dq, $J=2$, 7 Hz, 18-H), 4.22 (1H, dt, $J=8$, 2 Hz, 17-H), 4.00–4.09 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 3.62–3.67 (2H, m, $COOCH_2CH_2OH$), 2.22–2.59 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=644$. Calcd for $C_{34}H_{36}N_4O_5Zn$: M^+ , 644.

4.4.12. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a diethylene glycol monoester (2b). Zinc metallation of **7b** gave dark green solids of **2b**, 90% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 9.3 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 601 (0.10), 566 (0.06), 523 (0.04), 424 (1.00), 404 nm (0.56); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.56$, 9.31, 8.35 (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.20, 5.07 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.41 (1H, dq, $J=2$, 7 Hz, 18-H), 4.21 (1H, dt, $J=8$, 2 Hz, 17-H), 4.06–4.16 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 3.64–3.67, 3.45–3.58 (2H+4H, m, $COOCH_2CH_2OCH_2CH_2OH$), 2.22–2.64 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.70 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=688$. Calcd for $C_{36}H_{40}N_4O_6Zn$: M^+ , 688.

4.4.13. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a triethylene glycol monoester (2c). Zinc metallation of **7c** gave dark green solids of **2c**, 89% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 9.2 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.74), 602 (0.10), 568 (0.06), 521 (0.04), 424 (1.00), 405 nm (0.59); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57$, 9.32, 8.36 (each 1H, s, 5-, 10-, 20-H), 5.85, 5.81 (each 1H, d, $J=13$ Hz, 3- CH_2), 5.18, 5.06 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.40 (1H, dq, $J=2$, 7 Hz, 18-H), 4.22 (1H, dt, $J=8$, 2 Hz, 17-H), 3.96–4.09 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12-

CH_3), 3.36–3.53 (10H, m, $COOCH_2CH_2O(CH_2CH_2O)_2H$), 2.20–2.59 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=732$. Calcd for $C_{38}H_{44}N_4O_7Zn$: M^+ , 732.

4.4.14. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a tetraethylene glycol monoester (2d). Zinc metallation of **7d** gave dark green solids of **2d**, 93% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 9.1 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.73), 603 (0.10), 567 (0.05), 521 (0.03), 424 (1.00), 405 nm (0.56); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57$, 9.32, 8.36 (each 1H, s, 5-, 10-, 20-H), 5.84, 5.81 (each 1H, d, $J=13$ Hz, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.40 (1H, dq, $J=2$, 7 Hz, 18-H), 4.22 (1H, dt, $J=8$, 2 Hz, 17-H), 3.93–4.09 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 3.33–3.51 (14H, m, $COOCH_2CH_2O(CH_2CH_2O)_3H$), 2.26–2.60 (4H, m, 17- CH_2CH_2), 1.71 (3H, d, $J=7$ Hz, 18- CH_3), 1.70 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=776$. Calcd for $C_{40}H_{48}N_4O_8Zn$: M^+ , 776.

4.4.15. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 300 monoester (2e). Zinc metallation of **7e** gave dark green solids of **2e**, 92% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 8.3 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.73), 602 (0.10), 566 (0.05), 521 (0.03), 424 (1.00), 405 nm (0.56); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57$, 9.33, 8.36 (each 1H, s, 5-, 10-, 20-H), 5.85, 5.81 (each 1H, d, $J=13$ Hz, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.40 (1H, dq, $J=2$, 7 Hz, 18-H), 4.22 (1H, dt, $J=8$, 2 Hz, 17-H), 3.96–4.13 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 3.33–3.68 (24H, m, $COOCH_2CH_2O(CH_2CH_2O)_{5.4}$), 2.24–2.59 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=732$, 776, 820, 864, 908, 952, 996, 1040. Calcd for $C_{38}H_{44}N_4O_7Zn$: M^+ , 732 ($n=3$), $C_{40}H_{48}N_4O_8Zn$: M^+ , 776 ($n=4$), $C_{42}H_{52}N_4O_9Zn$: M^+ , 820 ($n=5$), $C_{44}H_{56}N_4O_{10}Zn$: M^+ , 864 ($n=6$), $C_{46}H_{60}N_4O_{11}Zn$: M^+ , 908 ($n=7$), $C_{48}H_{64}N_4O_{12}Zn$: M^+ , 952 ($n=8$), $C_{50}H_{68}N_4O_{13}Zn$: M^+ , 996 ($n=9$), $C_{52}H_{72}N_4O_{14}Zn$: M^+ , 1040 ($n=10$).

4.4.16. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 400 monoester (2f). Zinc metallation of **7f** gave dark green solids of **2f**, 88% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 8.1 min). Mp $>300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 601 (0.10), 567 (0.05), 520 (0.03), 424 (1.00), 405 nm (0.56); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57$, 9.33, 8.36 (each 1H, s, 5-, 10-, 20-H), 5.84, 5.81 (each 1H, d, $J=13$ Hz, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13 2 - H_2), 4.40 (1H, dq, $J=2$, 7 Hz, 18-H), 4.22 (1H, dt, $J=8$, 2 Hz, 17-H), 4.00–4.14 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 3.36–3.69 (33H, m, $COOCH_2CH_2O(CH_2CH_2O)_{7.7}$), 2.25–2.60 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8 1 - CH_3). MS (FAB) found: $m/z=776$, 820, 864, 908, 952, 996, 1040, 1084, 1128. Calcd for $C_{40}H_{48}N_4O_8Zn$: M^+ , 776 ($n=4$), $C_{42}H_{52}N_4O_9Zn$: M^+ , 820 ($n=5$), $C_{44}H_{56}N_4O_{10}Zn$: M^+ , 864

($n=6$), $C_{46}H_{60}N_4O_{11}Zn$: M^+ , 908 ($n=7$), $C_{48}H_{64}N_4O_{12}Zn$: M^+ , 952 ($n=8$), $C_{50}H_{68}N_4O_{13}Zn$: M^+ , 996 ($n=9$), $C_{52}H_{72}N_4O_{14}Zn$: M^+ , 1040 ($n=10$), $C_{54}H_{76}N_4O_{15}Zn$: M^+ , 1084 ($n=11$), $C_{56}H_{80}N_4O_{16}Zn$: M^+ , 1128 ($n=12$).

4.4.17. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 600 monoester (2g). Zinc metallation of **7g** gave dark green solids of **2g**, 93% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 7.8 min), $Mp >300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 602 (0.10), 567 (0.05), 522 (0.03), 424 (1.00), 405 nm (0.56); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57, 9.32, 8.36$ (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13^2-H_2), 4.39 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 4.00–4.13 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.39–3.73 (51H, m, $COOCH_2CH_2O(CH_2CH_2O)_{12.3}$), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 2.23–2.61 (4H, m, 17- CH_2CH_2), 1.71 (3H, d, $J=7$ Hz, 18- CH_3), 1.70 (3H, t, $J=7$ Hz, 8¹- CH_3). MS (FAB) found: $m/z=908, 952, 996, 1040, 1084, 1128, 1172, 1216, 1260, 1304, 1348$. Calcd for $C_{46}H_{60}N_4O_{11}Zn$: M^+ , 908 ($n=7$), $C_{48}H_{64}N_4O_{12}Zn$: M^+ , 952 ($n=8$), $C_{50}H_{68}N_4O_{13}Zn$: M^+ , 996 ($n=9$), $C_{52}H_{72}N_4O_{14}Zn$: M^+ , 1040 ($n=10$), $C_{54}H_{76}N_4O_{15}Zn$: M^+ , 1084 ($n=11$), $C_{56}H_{80}N_4O_{16}Zn$: M^+ , 1128 ($n=12$), $C_{58}H_{84}N_4O_{17}Zn$: M^+ , 1172 ($n=13$), $C_{60}H_{88}N_4O_{18}Zn$: M^+ , 1216 ($n=14$), $C_{62}H_{92}N_4O_{19}Zn$: M^+ , 1260 ($n=15$), $C_{64}H_{96}N_4O_{20}Zn$: M^+ , 1304 ($n=16$), $C_{66}H_{100}N_4O_{21}Zn$: M^+ , 1348 ($n=17$).

4.4.18. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 1000 monoester (2h). Zinc metallation of **7h** gave dark green solids of **2h**, 87% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 7.4 min), $Mp >300^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 602 (0.10), 566 (0.05), 521 (0.03), 424 (1.00), 405 nm (0.55); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57, 9.33, 8.36$ (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13^2-H_2), 4.40 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 4.00–4.14 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.41–3.73 (88H, m, $COOCH_2CH_2O(CH_2CH_2O)_{21.6}$), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 2.20–2.61 (4H, m, 17- CH_2CH_2), 1.71 (3H, d, $J=7$ Hz, 18- CH_3), 1.70 (3H, t, $J=7$ Hz, 8¹- CH_3). MS (FAB) found: $m/z=1172, 1216, 1260, 1304, 1348, 1392, 1436, 1480, 1524, 1568, 1612, 1656, 1700, 1744, 1788, 1832, 1876, 1920$. Calcd for $C_{58}H_{84}N_4O_{17}Zn$: M^+ , 1172 ($n=13$), $C_{60}H_{88}N_4O_{18}Zn$: M^+ , 1216 ($n=14$), $C_{62}H_{92}N_4O_{19}Zn$: M^+ , 1260 ($n=15$), $C_{64}H_{96}N_4O_{20}Zn$: M^+ , 1304 ($n=16$), $C_{66}H_{100}N_4O_{21}Zn$: M^+ , 1348 ($n=17$), $C_{68}H_{104}N_4O_{22}Zn$: M^+ , 1392 ($n=18$), $C_{70}H_{108}N_4O_{23}Zn$: M^+ , 1436 ($n=19$), $C_{72}H_{112}N_4O_{24}Zn$: M^+ , 1480 ($n=20$), $C_{74}H_{116}N_4O_{25}Zn$: M^+ , 1524 ($n=21$), $C_{76}H_{120}N_4O_{26}Zn$: M^+ , 1568 ($n=22$), $C_{78}H_{124}N_4O_{27}Zn$: M^+ , 1612 ($n=23$), $C_{80}H_{128}N_4O_{28}Zn$: M^+ , 1656 ($n=24$), $C_{82}H_{132}N_4O_{29}Zn$: M^+ , 1700 ($n=25$), $C_{84}H_{136}N_4O_{30}Zn$: M^+ , 1744 ($n=26$), $C_{86}H_{140}N_4O_{31}Zn$: M^+ , 1788 ($n=27$), $C_{88}H_{144}N_4O_{32}Zn$: M^+ , 1832 ($n=28$), $C_{90}H_{148}N_4O_{33}Zn$: M^+ , 1876 ($n=29$), $C_{92}H_{152}N_4O_{34}Zn$: M^+ , 1920 ($n=30$).

4.4.19. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 1540 monoester (2i). Zinc metallation of **7i** gave dark green solids of **2i**, 89% yield

(HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 6.9 min). Mp 221–224 $^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.75), 603 (0.10), 566 (0.05), 520 (0.03), 424 (1.00), 404 nm (0.55); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57, 9.33, 8.36$ (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13^2-H_2), 4.40 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 4.01–4.14 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.41–3.83 (134H, m, $COOCH_2CH_2O(CH_2CH_2O)_{33}$), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 2.25–2.60 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8¹- CH_3). MS (FAB) found: $m/z=1656, 1700, 1744, 1788, 1832, 1876, 1920, 1964, 2008, 2052, 2096, 2140, 2184, 2228, 2272, 2316, 2360, 2404, 2448$. Calcd for $C_{80}H_{128}N_4O_{28}Zn$: M^+ , 1656 ($n=24$), $C_{82}H_{132}N_4O_{29}Zn$: M^+ , 1700 ($n=25$), $C_{84}H_{136}N_4O_{30}Zn$: M^+ , 1744 ($n=26$), $C_{86}H_{140}N_4O_{31}Zn$: M^+ , 1788 ($n=27$), $C_{88}H_{144}N_4O_{32}Zn$: M^+ , 1832 ($n=28$), $C_{90}H_{148}N_4O_{33}Zn$: M^+ , 1876 ($n=29$), $C_{92}H_{152}N_4O_{34}Zn$: M^+ , 1920 ($n=30$), $C_{94}H_{156}N_4O_{35}Zn$: M^+ , 1964 ($n=31$), $C_{96}H_{160}N_4O_{36}Zn$: M^+ , 2008 ($n=32$), $C_{98}H_{164}N_4O_{37}Zn$: M^+ , 2052 ($n=33$), $C_{100}H_{168}N_4O_{38}Zn$: M^+ , 2096 ($n=34$), $C_{102}H_{172}N_4O_{39}Zn$: M^+ , 2140 ($n=35$), $C_{104}H_{176}N_4O_{40}Zn$: M^+ , 2184 ($n=36$), $C_{106}H_{180}N_4O_{41}Zn$: M^+ , 2228 ($n=37$), $C_{108}H_{184}N_4O_{42}Zn$: M^+ , 2272 ($n=38$), $C_{110}H_{188}N_4O_{43}Zn$: M^+ , 2316 ($n=39$), $C_{112}H_{192}N_4O_{44}Zn$: M^+ , 2360 ($n=40$), $C_{114}H_{196}N_4O_{45}Zn$: M^+ , 2404 ($n=41$), $C_{116}H_{200}N_4O_{46}Zn$: M^+ , 2448 ($n=42$).

4.4.20. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a polyethylene glycol 2000 monoester (2j). Zinc metallation of **7j** gave dark green solids of **2j**, 84% yield (HPLC, Cosmosil with 10% $H_2O/MeOH$ 1.0 mL/min, the retention time was 6.6 min), Mp 208–212 $^\circ C$; VIS (THF) $\lambda_{max}=647$ (0.76), 600 (0.10), 568 (0.05), 522 (0.03), 424 (1.00), 404 nm (0.55); 1H NMR (0.2% $C_5D_5N/CDCl_3$) $\delta=9.57, 9.33, 8.36$ (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3- CH_2), 5.17, 5.06 (each 1H, d, $J=20$ Hz, 13^2-H_2), 4.40 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 4.01–4.13 (2H, m, $COOCH_2$), 3.76 (2H, q, $J=7$ Hz, 8- CH_2), 3.41–3.83 (182H, m, $COOCH_2CH_2O(CH_2CH_2O)_{43.1}$), 3.69, 3.35, 3.26 (each 3H, s, 2-, 7-, 12- CH_3), 2.23–2.60 (4H, m, 17- CH_2CH_2), 1.72 (3H, d, $J=7$ Hz, 18- CH_3), 1.71 (3H, t, $J=7$ Hz, 8¹- CH_3). MS (FAB) found: $m/z=2008, 2052, 2096, 2140, 2184, 2228, 2272, 2316, 2360, 2404, 2448, 2492, 2536, 2580, 2624, 2668, 2712, 2756, 2800, 2844, 2888$. Calcd for $C_{96}H_{160}N_4O_{36}Zn$: M^+ , 2008 ($n=32$), $C_{98}H_{164}N_4O_{37}Zn$: M^+ , 2052 ($n=33$), $C_{100}H_{168}N_4O_{38}Zn$: M^+ , 2096 ($n=34$), $C_{102}H_{172}N_4O_{39}Zn$: M^+ , 2140 ($n=35$), $C_{104}H_{176}N_4O_{40}Zn$: M^+ , 2184 ($n=36$), $C_{106}H_{180}N_4O_{41}Zn$: M^+ , 2228 ($n=37$), $C_{108}H_{184}N_4O_{42}Zn$: M^+ , 2272 ($n=38$), $C_{110}H_{188}N_4O_{43}Zn$: M^+ , 2316 ($n=39$), $C_{112}H_{192}N_4O_{44}Zn$: M^+ , 2360 ($n=40$), $C_{114}H_{196}N_4O_{45}Zn$: M^+ , 2404 ($n=41$), $C_{116}H_{200}N_4O_{46}Zn$: M^+ , 2448 ($n=42$), $C_{118}H_{204}N_4O_{47}Zn$: M^+ , 2492 ($n=43$), $C_{120}H_{208}N_4O_{48}Zn$: M^+ , 2536 ($n=44$), $C_{122}H_{212}N_4O_{49}Zn$: M^+ , 2580 ($n=45$), $C_{124}H_{216}N_4O_{50}Zn$: M^+ , 2624 ($n=46$), $C_{126}H_{220}N_4O_{51}Zn$: M^+ , 2668 ($n=47$), $C_{128}H_{224}N_4O_{52}Zn$: M^+ , 2712 ($n=48$), $C_{130}H_{228}N_4O_{53}Zn$: M^+ , 2756 ($n=49$), $C_{132}H_{232}N_4O_{54}Zn$: M^+ , 2800 ($n=50$), $C_{134}H_{236}N_4O_{55}Zn$: M^+ , 2844 ($n=51$), $C_{136}H_{240}N_4O_{56}Zn$: M^+ , 2888 ($n=52$).

4.4.21. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-a 2-sulfoethyl amide (8). Amidation of pyropheophorbide-*d*

with taurine followed by reduction of 3-formyl group gave black solids of **8**, 59% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 5.0 min). Mp >300°C; VIS (MeOH) λ_{\max} =660 (0.46), 604 (0.10), 536 (0.11), 504 (0.11), 406 nm (1.00); ^1H NMR ($\text{CDCl}_3/\text{CD}_3\text{OD}=2/1$) δ =9.48, 9.44, 8.60 (each 1H, s, 5-, 10-, 20-H), 5.83 (2H, s, 3-CH₂), 5.31, 5.11 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.49 (1H, dq, $J=2, 7$ Hz, 18-H), 4.29 (1H, dt, $J=8, 2$ Hz, 17-H), 3.69 (2H, q, $J=7$ Hz, 8-CH₂), 3.64, 3.42, 3.25 (each 3H, s, 2-, 7-, 12-CH₃), 3.58 (2H, t, $J=6.4$ Hz, CONHCH₂), 2.91 (2H, t, $J=6.4$ Hz, CONHCH₂CH₂), 2.63–2.77, 2.40–2.52, 2.16–2.33 (1H+1H+2H, m, 17-CH₂CH₂), 1.81 (3H, d, $J=7$ Hz, 18-CH₃), 1.70 (3H, t, $J=7$ Hz, 8¹-CH₃), 0.14, –1.58 (each 1H, s, NH). MS (FAB) found: m/z =646. Calcd for C₃₄H₄₀N₅O₆S: MH⁺, 646.

4.4.22. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a 2-sulfoethyl amide sodium salt (3). Zinc metallation of **8** gave dark green solids of **3**, 88% yield (HPLC, Cosmosil with 25% H₂O/MeOH 1.0 mL/min, the retention time was 9.5 min). Mp >300°C; VIS (MeOH) λ_{\max} =653 (0.87), 607 (0.17), 575 (0.09), 427 (1.00), 408 nm (0.78); ^1H NMR ($\text{CDCl}_3/\text{CD}_3\text{OD}=2/1$) δ =9.49, 9.26, 8.34 (each 1H, s, 5-, 10-, 20-H), 5.76, 5.72 (each 1H, d, $J=13$ Hz, 3-CH₂), 5.23, 5.06 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.40 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 3.74 (2H, q, $J=7$ Hz, 8-CH₂), 3.63, 3.30, 3.26 (each 3H, s, 2-, 7-, 12-CH₃), 3.43–3.49 (2H, m, CONHCH₂), 2.79 (2H, t, $J=6$ Hz, CONHCH₂CH₂), 2.58–2.68, 2.01–2.43, (1H+3H, m, 17-CH₂CH₂), 1.78 (3H, d, $J=7$ Hz, 18-CH₃), 1.71 (3H, t, $J=7$ Hz, 8¹-CH₃). MS (FAB) found: m/z =752. Calcd for C₃₄H₃₆N₅O₆SZnNa₂: (M–H+Na)⁺, 752.

4.4.23. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-a 3-N,N-dimethylaminopropyl amide (9). Amidation of pyropheophorbide-d with *N,N*-dimethyl-1,3-propanediamine followed by reduction of 3-formyl group gave black solids of **9**, 63% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 8.8 min). Mp >300°C; VIS (CH₂Cl₂) λ_{\max} =662 (0.42), 605 (0.07), 536 (0.08), 504 (0.09), 410 nm (1.00); ^1H NMR (CDCl_3) δ =9.44, 9.39, 8.52 (each 1H, s, 5-, 10-, 20-H), 5.98–6.10 (1H, m, 17-CONH), 5.85, 5.79 (each 1H, d, $J=13$ Hz, 3-CH₂), 5.17, 5.05 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.49 (1H, dq, $J=2, 7$ Hz, 18-H), 4.29 (1H, dt, $J=8, 2$ Hz, 17-H), 3.67 (2H, q, $J=7$ Hz, 8-CH₂), 3.62, 3.38, 3.22 (each 3H, s, 2-, 7-, 12-CH₃), 2.88–3.17 (2H, m, CONHCH₂), 2.53–2.73, 2.25–2.44, 2.04–2.20 (4H, m, 17-CH₂CH₂), 1.76 (3H, d, $J=7$ Hz, 18-CH₃), 1.68 (3H, t, $J=7$ Hz, 8¹-CH₃), 1.08–1.24 (2H, m, CONHCH₂CH₂), 0.27, –1.81 (each 1H, s, NH). MS (FAB) found: m/z =623. Calcd for C₃₇H₄₇N₆O₃: MH⁺, 623.

4.4.24. 3-Devinyl-3-hydroxymethyl-pyropheophorbide-a 3-N,N,N-trimethylamoniopropyl amide iodide (6). *N*-methylation of **9** gave black solids of **6**, 72% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 4.8 min). Mp >300°C; VIS (MeOH) λ_{\max} =659 (0.48), 603 (0.09), 535 (0.09), 503 (0.09), 405 nm (1.00); ^1H NMR ($\text{CDCl}_3/\text{CD}_3\text{OD}=2/1$) δ =9.37, 9.36, 8.59 (each 1H, s, 5-, 10-, 20-H), 7.27 (1H, br, 17-CONH), 5.77, 5.72 (each 1H, d, $J=13$ Hz, 3-CH₂), 5.23, 5.03 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.55 (1H, dq, $J=2, 7$ Hz, 18-H), 4.24 (1H, dt, $J=8, 2$ Hz, 17-H), 3.64 (2H, q, $J=7$ Hz, 8-CH₂), 3.54, 3.37, 3.22

(each 3H, s, 2-, 7-, 12-CH₃), 3.09–3.18 (2H, m, CONHCH₂CH₂CH₂), 2.81–2.99 (2H, m, CONHCH₂), 2.50–2.61, 2.39–2.50, 2.12–2.32 (1H+1H+2H, m, 17-CH₂CH₂), 1.77 (3H, d, $J=7$ Hz, 18-CH₃), 1.66 (3H, t, $J=7$ Hz, 8¹-CH₃), 1.41–1.56 (2H, m, CONHCH₂CH₂), 0.88, –1.80 (each 1H, s, NH). MS (FAB) found: m/z =637. Calcd for C₃₈H₄₉N₆O₃: [M–I]⁺, 637.

4.4.25. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a 3-N,N-dimethylamino-propyl amide (10). Zinc metallation of **9** gave dark green solids of **10**, 89% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 3.5 min). Mp >300°C; VIS (CH₂Cl₂) λ_{\max} =654 (0.64), 612 (0.10), 575 (0.06), 524 (0.03), 428 nm (1.00); ^1H NMR ($\text{CDCl}_3/\text{CD}_3\text{OD}=2/1$) δ =9.56, 9.34, 8.34 (each 1H, s, 5-, 10-, 20-H), 5.84 (2H, s, 3-CH₂), 5.18, 5.05 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.45 (1H, dq, $J=2, 7$ Hz, 18-H), 4.21 (1H, dt, $J=8, 2$ Hz, 17-H), 3.75 (2H, q, $J=7$ Hz, 8-CH₂), 3.68, 3.32, 3.23 (each 3H, s, 2-, 7-, 12-CH₃), 2.86–3.16 (2H, m, CONHCH₂), 2.52–2.71, 2.23–2.41, 2.02–2.16 (4H, m, 17-CH₂CH₂), 2.16 (6H, s, N(CH₃)₂), 1.71 (3H, d, $J=7$ Hz, 18-CH₃), 1.69 (3H, t, $J=7$ Hz, 8¹-CH₃), 1.07–1.22 (2H, m, CONHCH₂CH₂). MS (FAB) found: m/z =685. Calcd for C₃₇H₄₅N₆O₃Zn: MH⁺, 685.

4.4.26. Zinc 3-devinyl-3-hydroxymethyl-pyropheophorbide-a 3-N,N,N-trimethyl-amoniopropyl amide iodide (4). *N*-Methylation of **10** gave dark green solids of **4**, 75% yield (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 4.4 min). Mp >300°C; VIS (MeOH) λ_{\max} =652 (0.69), 609 (0.16), 571 (0.08), 426 nm (1.00); ^1H NMR ($\text{CDCl}_3/\text{CD}_3\text{OD}=2/1$) δ =9.46, 9.21, 8.34 (each 1H, s, 5-, 10-, 20-H), 7.27 (1H, m, 17-CONH), 5.71, 5.66 (each 1H, d, $J=13$ Hz, 3-CH₂), 5.18, 5.04 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.40 (1H, dq, $J=2, 7$ Hz, 18-H), 4.22 (1H, dt, $J=8, 2$ Hz, 17-H), 3.70 (2H, q, $J=7$ Hz, 8-CH₂), 3.60, 3.27, 3.23 (each 3H, s, 2-, 7-, 12-CH₃), 2.95–3.02 (2H, m, CONHCH₂), 2.87 (2H, t, $J=6.4$ Hz, CONHCH₂CH₂CH₂N(CH₃)₃), 2.80 (9H, s, N(CH₃)₃), 2.55–2.65, 2.36–2.47, 2.18–2.29, 1.97–2.09 (each 1H, m, 17-CH₂CH₂), 1.78 (3H, d, $J=7$ Hz, 18-CH₃), 1.69 (3H, t, $J=7$ Hz, 8¹-CH₃), 1.37–1.49 (2H, m, CONHCH₂CH₂). MS (FAB) found: m/z =699. Calcd for C₃₈H₄₇N₆O₃Zn: [M–I]⁺, 699.

4.4.27. Zinc mesopyropheophorbide-a 3-N,N,N-trimethylamoniopropyl iodide (5). Amidation of mesopyropheophorbide-a (the 3-ethyl chlorin) with *N,N*-dimethyl-1,3-propanediamine followed by zinc metallation and *N*-methylation gave dark green solids of **5** (HPLC, Gelpack with MeOH 1.0 mL/min, the retention time was 5.2 min). Mp >300°C; VIS (MeOH) λ_{\max} =647 (0.81), 602 (0.18), 571 (0.10), 424 nm (1.00); ^1H NMR (0.4% C₅D₅N/CDCl₃) δ =9.46, 9.05, 8.26 (each 1H, s, 5-, 10-, 20-H), 6.55 (1H, m, 17-CONH), 5.20, 5.00 (each 1H, d, $J=20$ Hz, 13²-H₂), 4.42 (1H, dq, $J=2, 7$ Hz, 18-H), 4.15 (1H, dt, $J=8, 2$ Hz, 17-H), 3.71 (4H, q, $J=7$ Hz, 3-, 8-CH₂), 3.62, 3.23, 3.19 (each 3H, s, 2-, 7-, 12-CH₃), 3.23–3.29 (2H, m, CONHCH₂), 3.16 (2H, t, $J=6.4$ Hz, CONHCH₂CH₂CH₂N(CH₃)₃), 2.32 (9H, s, N(CH₃)₃), 2.05–2.40 (4H, m, 17-CH₂CH₂), 1.69 (3H, d, $J=7$ Hz, 18-CH₃), 1.673, 1.667 (each 3H, t, $J=7$ Hz, 3¹-, 8¹-CH₃), 1.53–1.66 (2H, m, CONHCH₂CH₂). MS (FAB) found: m/z =697. Calcd for C₃₉H₄₉N₆O₂Zn: [M–I]⁺, 697.

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References

1. (a) McDermott, G.; Prince, S. M.; Freer, A. A.; Lawless, H.-A. M.; Papiz, M. Z.; Cogdell, R. J. *Nature* **1995**, *374*, 517–521. (b) In *Photosynthesis: Photobiochemistry and Photobiophysics*; Ke, B., Ed.; Kluwer Academic: Dordrecht, 2001; pp 65–85.
2. (a) Olson, J. M. *Photochem. Photobiol.* **1998**, *67*, 61–75. (b) Tamiaki, H. *Coord. Chem. Rev.* **1996**, *148*, 183–197.
3. Holzwarth, A. R.; Schaffner, K. *Photosynth. Res.* **1994**, *41*, 225–233.
4. (a) Nozawa, T.; Ohtomo, K.; Suzuki, M.; Nakagawa, H.; Shikama, Y.; Konami, H.; Wang, Z.-Y. *Photosynth. Res.* **1994**, *41*, 211–223. (b) Matsuura, K.; Hirota, M.; Shimada, K.; Mimuro, M. *Photochem. Photobiol.* **1993**, *57*, 92–97.
5. Müller, M. G.; Griebenow, K.; Holzwarth, A. R. *Biochim. Biophys. Acta* **1993**, *1144*, 161–169.
6. Hildebrandt, P.; Tamiaki, H.; Holzwarth, A. R.; Schaffner, K. *J. Phys. Chem.* **1994**, *98*, 2192–2197.
7. (a) Mizoguchi, T.; Hara, K.; Nagae, H.; Koyama, Y. *Photochem. Photobiol.* **2000**, *71*, 596–609. (b) Ishii, T.; Kimura, M.; Yamamoto, T.; Kirihata, M.; Uehara, K. *Photochem. Photobiol.* **2000**, *71*, 567–573. (c) Steensgaard, D. B.; Wackerbarth, H.; Hildebrandt, P.; Holzwarth, A. R. *J. Phys. Chem. B* **2000**, *104*, 10379–10386. (d) Balaban, T. S.; Leitich, J.; Holzwarth, A. R.; Schaffner, K. *J. Phys. Chem. B* **2000**, *104*, 1362–1372. (e) Saga, Y.; Matsuura, K.; Tamiaki, H. *Photochem. Photobiol.* **2001**, *74*, 72–80. (f) Mizoguchi, T.; Saga, Y.; Tamiaki, H. *Photochem. Photobiol. Sci.* **2002**, *1*, 780–787.
8. Umetsu, M.; Seki, R.; Wang, Z.-Y.; Kumagai, I.; Nozawa, T. *J. Phys. Chem. B* **2002**, *106*, 3987–3995.
9. (a) Balaban, T. S.; Holzwarth, A. R.; Schaffner, K. *J. Mol. Struct.* **1995**, *349*, 183–186. (b) Griebenow, K.; Holzwarth, A. R.; van Mourik, F.; van Grondelle, R. van Mourik. *Biochim. Biophys. Acta.* **1991**, *1058*, 194–202.
10. Fages, F.; Griebenow, N.; Griebenow, K.; Holzwarth, A. R.; Schaffner, K. *J. Chem. Soc., Perkin Trans. 1* **1990**, 2791–2797.
11. Tamiaki, H.; Amakawa, M.; Shimono, Y.; Tanikaga, R.; Holzwarth, A. R.; Schaffner, K. *Photochem. Photobiol.* **1996**, *63*, 92–99.
12. Miyatake, T.; Tamiaki, H.; Holzwarth, A. R.; Schaffner, K. *Photochem. Photobiol.* **1999**, *69*, 448–456.
13. (a) Miyatake, T.; Tamiaki, H.; Holzwarth, A. R.; Schaffner, K. *Helv. Chim. Acta* **1999**, *82*, 797–810. (b) Prokhorenko, V. I.; Holzwarth, A. R.; Müller, M. G.; Schaffner, K.; Miyatake, T.; Tamiaki, H. *J. Phys. Chem. B* **2002**, *106*, 5761–5768.
14. (a) Yagai, S.; Miyatake, T.; Tamiaki, H. *J. Org. Chem.* **2002**, *67*, 49–58. (b) Yagai, S.; Miyatake, T.; Shimono, Y.; Tamiaki, H. *Photochem. Photobiol.* **2001**, *73*, 153–163. (c) Oba, T.; Tamiaki, H. *Supramol. Chem.* **2001**, *12*, 369–378. (d) Yagai, S.; Miyatake, T.; Tamiaki, H. *J. Photochem. Photobiol., B: Biol.* **1999**, *52*, 74–85. (e) Tamiaki, H.; Omoda, M.; Kubo, M. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1631–1632. (f) Tamiaki, H.; Tomida, T.; Miyatake, T. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1415–1418.
15. Tamiaki, H.; Holzwarth, A. R.; Schaffner, K. *Photosynth. Res.* **1994**, *41*, 245–251.
16. Miyatake, T.; Oba, T.; Tamiaki, H. *ChemBioChem* **2001**, *2*, 330–342.
17. Fuhrhop, J.-H. *Comprehensive Supramolecular Chemistry*; Lehn, J.-M., Atwood, J. L., Davies, J. E. D., Macnicol, D. D., Vögtle, F., Eds.; 1996; Vol. 9, pp 407–450.
18. Fabiano, A.-S.; Allouche, D.; Sanejouand, Y.-H.; Paillous, N. *Photochem. Photobiol.* **1997**, *66*, 336–345.
19. Tamiaki, H.; Miyata, S.; Kureishi, Y.; Tanikaga, R. *Tetrahedron* **1996**, *52*, 12421–12432.
20. Eichwurz, I.; Stiel, H.; Röder, B. *J. Photochem. Photobiol. B: Biol.* **2000**, *54*, 194–200.